



CROSS - LINKED ENZYME AGGREGATES OF RECOMBINANT XYLANASE
FOR CONVERSION OF HEMICELLULOSES TO XYLOOLIGOSACCHARIDES


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A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (*Bioprocess Engineering*)

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MARCH 2018

I declare that this thesis entitled “*Cross-linked Enzyme Aggregates of Recombinant Xylanase for Conversion of Hemicelluloses to Xylooligosaccharides*” is the result of my own research except as cited in the references. The thesis has not been accepted for any degree and is not concurrently submitted in candidature of any other degree.

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To my beloved husband (Muhamad Azraf bin Johana), mother (Hjh. Faridah Osman), brother (Prof. Dr. Sharifudin Md. Shaarani) and son (Haziq Naquiuddin).

ACKNOWLEDGEMENTS

Alhamdulillah. First and foremost, I owe my deepest gratitude to my supervisor, Prof. Dr. Rosli Md. Illias, whose encouragement, assistance, guidance and advices from the initial to the final stage, has greatly facilitated me in developing a good understanding of the subject. I wish to express my appreciation to my two co-supervisors, Prof. Dr. Jamaliah Md. Jahim (Universiti Kebangsaan Malaysia) and Prof. Madya Dr. Roshanida A. Rahman for the valuable discussions, great opinions and continuous motivation given. Special thanks go to the helpful laboratory staff, Mr. Yaakob, Mr. Latfi, Mr. Jefri and Madam Zainab for their kind assistance throughout this work. Without their help, the labwork might not have been completed successfully.

I am also greatly indebted to all my laboratory colleagues who have been helping me in numerous ways, particularly Dr. Nur Izyan, Dr. Hasmaliana, Dr. Fatimah, Dr. Baidurah, Dr. Rohaida, Izawati, Ling How Lie, Amalina, Aishah, Ummu Qistina, Rabiatul, Intan, Joyce, Samson, Hakimi, and Abbas.

Sincerely, I am thankful to the Ministry of Higher Education (MOHE) of Malaysia and Universiti Malaysia Pahang for funding my doctoral study as well as Universiti Teknologi Malaysia (UTM) for providing the facilities for my research.

Last but not least, my utmost appreciation to my beloved husband (Muhamad Azraf Johana), my loving mother (Hjh. Faridah Osman), my understanding brother (Prof. Dr. Sharifudin Md Shaarani), my son (Haziq Naquiuddin) and my cherished family members for their patience, eternal support and undying love during my studies.

ABSTRACT

Hemicelluloses are heterogeneous branched polymers of sugars that exist abundantly in nature. Enzymatic hydrolysis is envisioned as a highly potential method in converting hemicelluloses into fuels and value-added chemicals. However, the use of free enzyme is hampered by low operational stability, difficulty in recovery and non-reusability, which requires for enzyme immobilization. Carrier-bound immobilization leads to utilization of high cost matrices, clogging of filters during downstream processing and presence of large amounts of non-catalytic ballast. Therefore, cross-linked enzyme aggregates (CLEA), a carrier-free technology that combines purification (precipitation) and immobilization into a single operation and does not require purified enzymes, is the solution to these problems. In this study, a recombinant xylanase (Xyl) from *Trichoderma reesei* was immobilized using three approaches: Xyl-CLEA, Xyl-CLEA-BSA (bovine serum albumin) and Xyl-CLEA-silanized maghemite. The use of ethanol as precipitant (1:9 volume ratio of enzyme to precipitant), glutaraldehyde (0.2:1 of glutaraldehyde to enzyme of 100 mM concentration) as cross-linking agent and the introduction of (3-aminopropyl) triethoxysilane (APTES) silanized maghemite (0.0075:1 of silanized maghemite to enzyme) prevailed in forming xylanase CLEAs with good enzyme activity recovery (78 %), thermal stability (50 % retained activity) and reusability (50 % retained activity). The Xyl-CLEA-silanized maghemite enhanced the activity recovery 1.66- and 1.50-fold compared to Xyl-CLEA and Xyl-CLEA-BSA, respectively. At elevated temperature of 60 °C and pHs of 3.0 and 8.0, Xyl-CLEA-silanized maghemite achieved better stability compared to the other CLEAs and free enzyme. Xyl-CLEA-silanized maghemite also successfully retained more than 50 % of its activity after 6 cycles, whereas Xyl-CLEA only retained approximately 10 % after 5 cycles. Therefore, the performance of Xyl-CLEA-silanized maghemite was further investigated by xylan hydrolysis under optimised reaction conditions. Xylooligosaccharides yield was slightly improved by 1.26- fold compared to the free enzyme. Kinetic parameters confirmed that CLEA immobilization did affect the productivity of the designed biocatalyst.

ABSTRAK

Hemiselulosa adalah polimer bercabang heterogen yang wujud dengan sangat banyak dalam alam semulajadi. Hidrolisis enzimatik dibayangkan sebagai kaedah yang berpotensi tinggi untuk menukarkan hemiselulosa menjadi bahan api dan bahan kimia bernilai tambah. Walau bagaimanapun, penggunaan enzim bebas terhad disebabkan oleh kestabilan operasinya yang rendah, kesukaran untuk pemulihan dan ketidakbolehan guna semula, yang menyebabkan keperluan kepada proses imobilisasi enzim. Imobilisasi pembawa melibatkan penggunaan matriks yang berkos tinggi, penyumbatan penapis semasa pemprosesan hiliran dan kehadiran sejumlah besar balast bukan pemangkin. Oleh itu, agregat enzim terpaut silang (CLEA), teknologi bebas pembawa yang menggabungkan pemurnian (pemendakan) dan imobilisasi ke dalam satu operasi dan tidak memerlukan enzim tulen, adalah penyelesaian kepada masalah ini. Dalam kajian ini, *xylanase* rekombinan (Xyl) dari *Trichoderma reesei* diimobilisasi menggunakan tiga pendekatan: Xyl-CLEA, Xyl-CLEA-BSA (*bovine serum albumin*) dan Xyl-CLEA-*silanized maghemite*. Penggunaan etanol sebagai pemendak (1:9 nisbah isipadu enzim kepada pemendak), glutaraldehid (0.2:1 glutaraldehid kepada enzim berkepekatan 100 mM) sebagai agen pemaat silang dan pengenalan (*3-aminopropyl*) *triethoxysilane* (APTES) *silanized maghemite* (0.0075: 1 *silanized maghemite* kepada enzim) berjaya membentuk *xylanase* CLEA dengan pemulihan aktiviti enzim yang baik (78 %), kestabilan terma (50 % aktiviti tersimpan) dan kebolehan guna semula (50 % aktiviti tersimpan). Xyl-CLEA-*silanized maghemite* meningkatkan pemulihan aktiviti 1.66- dan 1.50- kali ganda berbanding dengan Xyl-CLEA dan Xyl-CLEA-BSA, masing-masing. Pada suhu tinggi 60 °C dan pH 3.0 dan 8.0, Xyl-CLEA-*silanized maghemite* mencapai kestabilan yang lebih baik berbanding dengan CLEA yang lain dan enzim bebas. Xyl-CLEA-*silanized maghemite* juga berjaya mengekalkan lebih daripada 50 % aktiviti selepas 6 kitaran, manakala Xyl-CLEA hanya mengekalkan kira-kira 10 % selepas 5 kitaran. Oleh itu, prestasi Xyl-CLEA-*silanized maghemite* dikaji selanjutnya melalui hidrolisis xilan di bawah keadaan tindak balas yang dioptimumkan. Hasil xilooligosakarida meningkat sebanyak 1.26- kali ganda berbanding dengan enzim bebas. Parameter kinetik mengesahkan bahawa imobilisasi CLEA mempengaruhi produktiviti biomangkin yang direka.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xv
	LIST OF SYMBOLS	xvii
	LIST OF APPENDICES	xviii
1	INTRODUCTION	1
	1.1 Introduction	1
	1.2 Objectives of Study	6
	1.3 Scopes of Study	7
	1.4 Rationale and Significance of the Study	8
2	LITERATURE REVIEW	10
	2.1 Lignocellulose	10
	2.2 Hemicellulases	14
	2.3 Xylanase (EC 3.2.1.8)	14

	2.4 Enzyme Stabilization	16
	2.5 Immobilization of Enzymes	19
	2.6 Enzyme Immobilization using Nanoparticles	22
	2.7 Cross-linked Enzyme Aggregates	24
	2.8 Research Gap	40
3	MATERIALS AND METHODS	41
	3.1 Overall Research Design	41
	3.2 Chemicals	43
	3.3 Enzyme Preparation	43
	3.4 Yeast Stock Preparation and Storage	44
	3.5 Yeast Culture and Expression Media	44
	3.6 Cultivation of Recombinant <i>Pichia pastoris</i>	45
	3.7 Scale-up Expression	45
	3.8 Enzyme Partial Purification Process	46
	3.9 Addition of PMSF	46
	3.10 Enzyme Immobilization via CLEA	47
	3.11 Enzyme Characterization	51
	3.12 Enzymatic Reaction	57
4	RESULTS AND DISCUSSIONS	65
	4.1 Introduction	65
	4.2 Preparation of Cross-linked Enzyme Aggregates	66
	4.3 Preparation of Xyl-CLEA-BSA	81
	4.4 Preparation of Xyl-CLEA-Silanized Maghemite	84
	4.5 Characterization of Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-Silanized Maghemite	89
	4.6 Reusability	97
	4.7 Characteristic Analysis of Silanized Maghemite and Xyl-CLEAs	98
	4.8 Xyl-CLEA-Silanized Maghemite in Xylan Hydrolysis	103

5	CONCLUSIONS AND RECOMMENDATIONS	142
5.1	Conclusions	142
5.2	Recommendations for Future Work	143
	REFERENCES	146
	Appendices A - J	166 - 194

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Advantages and disadvantages of various enzyme stabilization techniques	17
2.2	Potential of different enzyme immobilization strategies in improving enzyme performance	21
2.3	Immobilization of enzymes on different nanomaterials	23
2.4	Findings on magnetic CLEAs	39
3.1	Details of the low and high limit for each parameter used in the Factorial screening design	62
3.2	Details of the low and high limit for each parameter used in the Central Composite Design	63
4.1	Infrared absorption bands associated with Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite, silanized and non-silanized maghemite	99
4.2	Characteristics of Xyl-CLEA-silanized maghemite and free xylanase	104
4.3	Chemical composition of beechwood xylan	106
4.4	Experimental design and results of full factorial design	120
4.5	Analysis of variance of the factorial design for reducing sugar production	121
4.6	Statistical analysis for reducing sugar production in xylan hydrolysis by Xyl-CLEA-silanized maghemite	122

4.7	Experimental design and results of the Central Composite Design	124
4.8	ANOVA for response surface quadratic model for reducing sugar yield	125
4.9	ANOVA for response surface linear model for XOS yield	126
4.10	Statistical analysis for reducing sugar yield and XOS yield in xylan hydrolysis by Xyl-CLEA-silanized maghemite	128
4.11	Summary of the optimized reaction conditions for the xylan hydrolysis using Xyl-CLEA-silanized maghemite	134
4.12	Kinetic analysis of xylanase on beechwood xylan	136

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Diagrammatic illustration of the lignocellulosic framework	12
2.2	General scheme for the production of cross-linked enzyme aggregates (CLEAs)	27
2.3	The SEM images of (a) sugar-free CLEAs; (b) 20.0% trehalose-assisted CLEAs	35
2.4	The SEM images of (a) CLEAs and (b) p-CLEAs	37
3.1	Overall research design	42
3.2	Double reciprocal (Lineweaver-Burk Plot) and kinetic parameters determination	64
4.1	Xylanase recovery in Xyl-CLEA preparation using different types of precipitants	69
4.2	Effect of different enzyme to precipitant ratios on the relative activity of Xyl-CLEA	71
4.3	Xylanase recovery in Xyl-CLEA preparation at different cross-linking times	72
4.4	Xylanase recovery in Xyl-CLEA preparation using varied concentrations of glutaraldehyde	74
4.5	Xylanase recovery in Xyl-CLEA preparation using different glutaraldehyde to enzyme ratios (v: v)	77
4.6	Effect of additives on the relative activity of Xyl-CLEA	79

4.7	Xylanase recovery in Xyl-CLEAs and percent enzyme loss in supernatant phase after cross-linking using different additives	80
4.8	Xylanase activity recovery in Xyl-CLEA-BSA using different amount of BSA	82
4.9	Xylanase recovery in Xyl-CLEAs and Xyl-CLEA-BSA using different amounts of enzyme loading	84
4.10	Effect of silanization of maghemite particles on Xyl-CLEAs.	87
4.11	Process of enzyme cross-linking using APTES modified nanoparticles	88
4.12	Effect of temperature on enzyme activity recovery of free enzyme, Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite	90
4.13	Temperature stabilities of free enzyme, Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite	92
4.14	Effect of pH on enzyme activity recovery of free enzyme, Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite	94
4.15	pH stabilities of free enzyme, Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite	96
4.16	Reusability of Xyl-CLEA, Xyl-CLEA-BSA and Xyl-CLEA-silanized maghemite	98
4.17	FTIR spectra of a) non-silanized maghemite and b) silanized maghemite	100
4.18	FTIR spectra of a) Xyl-CLEA-silanized maghemite, b) Xyl-CLEA-BSA and c) Xyl-CLEA	101
4.19	FESEM images of a) Xyl-CLEA, b) Xyl-CLEA-BSA and c) Xyl-CLEA-silanized maghemite	102
4.20	Structure of hardwood xylan	106
4.21	Effect of reaction volume on i) reducing sugar concentration ii) reducing sugar yield	108
4.22	Effect of reaction time on i) reducing sugar concentration ii) reducing sugar yield	109
4.23	Temperature stability of Xyl-CLEA-silanized maghemite.	110

4.24	Effect of enzyme dosage on i) reducing sugar concentration ii) reducing sugar yield.	112
4.25	Effect of temperature on i) reducing sugar concentration ii) reducing sugar yield	114
4.26	Effect of ph on i) reducing sugar concentration ii) reducing sugar yield	116
4.27	Effect of substrate concentration on i) reducing sugar concentration ii) reducing sugar yield	118
4.28	3D response surface plot of (a) reducing sugar yield (b) XOS yield	129
4.29	3D response surface plot of (a) reducing sugar yield (b) XOS yield	131
4.30	FESEM images of (a) Xyl-CLEA-silanized maghemite and (b) free xylanase	141

LIST OF ABBREVIATIONS

A	-	Absorbance
ANOVA	-	Analysis of variance
ATCC	-	American type culture collection
B	-	Blank / buffer only
BMGY	-	Buffered glycerol-complex medium
BMMY	-	Buffered methanol-complex medium
BSA	-	Bovine serum albumin
CCD	-	Central composite design
dH ₂ O	-	Distilled water
DNS	-	3, 5-Dinitrosalicylic acid
EC	-	Enzyme control
e.g.	-	“for example”
etc.	-	et cetera / “and so forth”
g	-	gram
g/L	-	gram per liter
h	-	hour
HCl	-	Hydrochloric acid
HPLC	-	High performance liquid chromatography
<i>i.e.</i>	-	“that is”
kDa	-	kilo Dalton
L	-	Liter
M	-	Molar
min	-	minute
mL	-	milliliter
mm	-	millimete

mM	-	milimolar
MWCO	-	Molecular weight cut off
NaOH	-	Sodium hydroxide
nm	-	nanometer
OD	-	Optical density
OFAT	-	One-factor-at-a time
pI	-	Isoelectric point
PMSF	-	Phenylmethysulfonyl fluoride
PEG	-	Polyethylene glycol
RBB	-	Remazol brilliant blue
RI	-	Refractive index
Rpm	-	revolutions per minute
RSM	-	Response surface methodology
SC	-	Substrate control
SD	-	Standard deviation
SDS PAGE	-	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
T	-	Temperature
TRS	-	Total reducing sugar
U	-	Unit enzyme
UV	-	Ultra violet
V	-	Velocity
v/v	-	Volume per volume
w/w	-	Weight per weight
w/v	-	Weight per volume
X1	-	Xylose
X2	-	Xylobiose
X3	-	Xylotriose
X4	-	Xylotetraose
X5	-	Xylopentaoase
X6	-	Xylohexaoase
XOS	-	Xylooligosaccharides
Xyn2	-	Xylanase2
YPD	-	Yeast extract peptone dextrose

LIST OF SYMBOLS

°C	-	Degree Celsius
°F	-	Degree Fahrenheit
%	-	Percentage
α	-	alpha
β	-	beta
γ	-	gamma
μL	-	microlitre
10X D	-	Ten times dilution
$\mu\text{g/mL}$	-	microgram per litre
1X	-	One time
μmol	-	micromole
5X	-	Five times
3D	-	Three dimensions
™	-	Trademark symbol
~	-	Approximate value

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Preparation of buffers, mediums and reagents	166
B	Mathematical equations	176
C	Graphs of standard curve	180
D	Nucleotide sequence and protein structure	181
E	Standard procedure for instruments	183
F	Pictures of FESEM analysis	185
G	SDS PAGE and RBB plate of Xyn2	187
H	HPLC analysis	189
I	Simplified illustration on formation of Xyl-CLEA-silanized maghemite	192
J	List of publications	193

REFERENCES

- Ahmed, S., Imdad, S. S., & Jamil, A. (2012). Comparative study for the kinetics of extracellular xylanases from *Trichoderma harzianum* and *Chaetomium thermophilum*. *Electronic Journal of Biotechnology*, 15(3).
- Akpinar, O., Erdogan, K., & Bostanci, S. (2009). Production of xylooligosaccharides by controlled acid hydrolysis of lignocellulosic materials. *Carbohydrate Research*, 344(5), 660–666.
- Alias, N. I., Mahadi, N. M., Munir, A., Murad, A., Abu, F. D., Rabu, A., & Illias, R. M. (2011). Expression optimisation of recombinant α -L- arabinofuranosidase from *Aspergillus niger* ATCC 120120 in *Pichia pastoris* and its biochemical characterisation, 10(35), 6700–6710.
- Alvarenga, A. E., Amoroso, M. J., Illanes, A., & Castro, G. R. (2014). Cross-linked α -l-rhamnosidase aggregates with potential application in food industry. *European Food Research and Technology*, 238(5), 797–801.
- Ansari, S. A., & Husain, Q. (2012). Potential applications of enzymes immobilized on/in nano materials: A review. *Biotechnology Advances*, 30(3), 512–23.
- Asco-Lozano, Sv., López-Gallego, F., Vázquez-Duhalt, R., Mateos-Díaz, J. C., Guisán, J. M., & Favela-Torres, E. (2014). Carrier-Free Immobilization of Lipase from *Candida rugosa* with Polyethyleneimines by Carboxyl-Activated Cross-Linking. *Biomacromolecules*, 15(5), 1896–903.
- Ashtari, K., Khajeh, K., Fasihi, J., Ashtari, P., Ramazani, A., & Vali, H. (2012). Silica-encapsulated magnetic nanoparticles: enzyme immobilization and cytotoxic study. *International Journal of Biological Macromolecules*, 50(4), 1063–9.
- Ayhan, H., Ayhan, F., & Gulsu, A. (2012). Highly biocompatible enzyme aggregates cross-linked by L-lysine. *Turkish Journal of Biochemistry*, 37(1), 14–20.
- Aytar, B. S., & Bakir, U. (2008). Preparation of cross-linked tyrosinase aggregates, *Process Biochemistry*, 43(2), 125–131.

- Bailey, M. J., Biely, P., & Poutanen, K. (1992). Interlaboratory testing of methods for assay of xylanase activity. *Journal of Biotechnology*, 23(3), 257–270.
- Barbosa, O., Ortiz, C., Berenguer-Murcia, Á., Torres, R., Rodrigues, R. C., & Fernandez-Lafuente, R. (2014). Glutaraldehyde in bio-catalysts design: a useful cross-linker and a versatile tool in enzyme immobilization. *RSC Advances*, 4(4), 1583.
- Bee, A., Massart, R., & Neveu, S. (1995). Synthesis of very fine maghemite particles. *Journal of Magnetism and Magnetic Materials*, 149(1–2), 6–9.
- Berrin, J.-G., & Juge, N. (2008). Factors affecting xylanase functionality in the degradation of arabinoxylans. *Biotechnology Letters*, 30(7), 1139–50.
- Bhattacharya, A., & Pletschke, B. I. (2014). Magnetic cross-linked enzyme aggregates (CLEAs): A novel concept towards carrier free immobilization of lignocellulolytic enzymes. *Enzyme and Microbial Technology*, 61–62, 17–27.
- Bhattacharya, A., & Pletschke, B. I. (2015). Strategic optimization of xylanase – mannanase combi-CLEAs for synergistic and efficient hydrolysis of complex lignocellulosic substrates. *Journal of Molecular Catalysis. B, Enzymatic*, 115, 140–150.
- Bian, J., Peng, F., Peng, X. P., Peng, P., Xu, F., & Sun, R. C. (2013). Structural features and antioxidant activity of xylooligosaccharides enzymatically produced from sugarcane bagasse. *Bioresource Technology*, 127, 236–241.
- Börjesson, M., & Westman, G. (2016). Branching of hemicelluloses through an azetidinium salt ring-opening reaction. *Carbohydrate Research*, 428, 23–30.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1–2), 248–254.
- Bruggink, A., Schoevaart, R., & Kieboom, T. (2003). Concepts of Nature in Organic Synthesis: Cascade Catalysis and Multistep Conversions in Concert. *Organic Process Research & Development*, 7(5), 622–640.
- Cabana, H., Jones, J. P., & Agathos, S. N. (2007). Preparation and characterization of cross-linked laccase aggregates and their application to the elimination of endocrine disrupting chemicals. *Journal of Biotechnology*, 132(1), 23–31.

- Cabana, H., Jones, J. P., & Agathos, S. N. (2009). Utilization of cross-linked laccase aggregates in a perfusion basket reactor for the continuous elimination of endocrine-disrupting chemicals. *Biotechnology and Bioengineering*, 102(6), 1582-1592.
- Čampelj, S., Makovec, D., & Drofenik, M. (2009). Functionalization of magnetic nanoparticles with 3-aminopropyl silane. *Journal of Magnetism and Magnetic Materials*, 321(10), 1346–1350.
- Cantarel, B. I., Coutinho, P. M., Rancurel, C., Bernard, T., Lombard, V., & Henrissat, B. (2009). The Carbohydrate-Active EnZymes database (CAZy): An expert resource for glycogenomics. *Nucleic Acids Research*, 37(SUPPL. 1), 233–238.
- Cao, L., Langen, L. Van, & Sheldon, R. a. (2003). Immobilised enzymes: carrier-bound or carrier-free? *Current Opinion in Biotechnology*, 14(4), 387–394.
- Cerdobbel, A., De Winter, K., Desmet, T., & Soetaert, W. (2010). Sucrose phosphorylase as cross-linked enzyme aggregate: improved thermal stability for industrial applications. *Biotechnology Journal*, 5(11), 1192–1197.
- Cha, T. W., Quo, A., & Zhu, X. Y. (2005). Enzymatic activity on a chip: The critical role of protein orientation. *Proteomics*, 5(2), 416–419.
- Chang, L., Shepherd, D., Sun, J., Ouellette, D., Grant, K. L., Tang, X., & Pikal, M. J. (2005). Mechanism of protein stabilization by sugars during freeze-drying and storage: Native structure preservation, specific interaction, and/or immobilization in a glassy matrix? *Journal of Pharmaceutical Sciences*, 94(7), 1427–1444.
- Chapla, D., Pandit, P., & Shah, A. (2012). Bioresource Technology Production of xylooligosaccharides from corncob xylan by fungal xylanase and their utilization by probiotics. *Bioresource Technology*, 115, 215–221.
- Chaplin, M. and Bucke, C. (1990). *Concentration by Precipitation, Enzyme Technology*, Cambridge University Press.
- Chaudhari, S. A., & Singhal, R. S. (2017). A strategic approach for direct recovery and stabilization of *Fusarium sp.* ICT SAC1 cutinase from solid state fermented broth by carrier free cross-linked enzyme aggregates. *International Journal of Biological Macromolecules*, 98, 610–621.
- Chen, Y., Xiao, C., Chen, X., Yang, L., Qi, X., Zheng, J., Li, M. & Zhang, J. (2014). Preparation of cross-linked enzyme aggregates in water-in-oil emulsion: Application to trehalose synthase. *Journal of Molecular Catalysis B: Enzymatic*, 100, 84–90.

- Chen, Z., Wang, Y., Liu, W., Wang, J., & Chen, H. (2017). A novel cross-linked enzyme aggregates (CLEAs) of papain and neutrase-production, partial characterization and application. *International Journal of Biological Macromolecules*, 95, 650–657.
- Chmura, A., Rustler, S., Paravidino, M., Rantwijk, F. Van, Stolz, A., & Sheldon, R. A. (2013). Tetrahedron: Asymmetry The combi-CLEA approach: enzymatic cascade synthesis of enantiomerically pure (S) -mandelic acid. *Tetrahedron: Asymmetry*, 24(19), 1225–1232.
- Cipolatti, E. P., Silva, M. J. a., Klein, M., Feddern, V., Feltes, M. M. C., Oliveira, J. V., Ninow, J. L. & de Oliveira, D. (2014). Current status and trends in enzymatic nanoimmobilization. *Journal of Molecular Catalysis B: Enzymatic*, 99, 56–67.
- Collins, T., Gerday, C., & Feller, G. (2005). Xylanases, xylanase families and extremophilic xylanases. *FEMS Microbiology Reviews*, 29(1), 3–23.
- Cortez, E. V., & Jr, A. P. (1999). Xylanase and b -xylosidase separation by fractional precipitation, 35, 277–283.
- Cosio-Castañeda, C., Martínez-García, R., & Socolovsky, L. M. (2014). Synthesis of silanized maghemite nanoparticles onto reduced graphene sheets composites. *Solid State Sciences*, 30, 17–20.
- Cowan, D. a, & Fernandez-Lafuente, R. (2011). Enhancing the functional properties of thermophilic enzymes by chemical modification and immobilization. *Enzyme and Microbial Technology*, 49(4), 326–46.
- Cui, J. D., Cui, L. L., Zhang, S. P., Zhang, Y. F., Su, Z. G., & Ma, G. H. (2014). Hybrid magnetic cross-linked enzyme aggregates of phenylalanine ammonia lyase from *Rhodotorula glutinis*. *PloS One*, 9(5), e97221.
- Cui, J. D., & Jia, S. R. (2015). Optimization protocols and improved strategies of cross-linked enzyme aggregates technology: current development and future challenges, 8551(October).
- Dalal, S., Kapoor, M., & Gupta, M. N. (2007). Preparation and characterization of combi-CLEAs catalyzing multiple non-cascade reactions. *Journal of Molecular Catalysis B: Enzymatic*, 44(3–4), 128–132.
- Dalal, S., Sharma, A., & Gupta, M. N. (2007). A multipurpose immobilized biocatalyst with pectinase, xylanase and cellulase activities. *Chemistry Central Journal*, 1, 16.

- Das, A., Singh, J., & Yogalakshmi, K. N. (2017). Laccase immobilized magnetic iron nanoparticles: Fabrication and its performance evaluation in chlorpyrifos degradation. *International Biodeterioration and Biodegradation*, 117, 183–189.
- Davis, B. G. (2003). Chemical modification of biocatalysts. *Current Opinion in Biotechnology*, 14(4), 379–386.
- De Vries, R. P., Kester, H. C. M., Poulsen, C. H., Benen, J. A. E., & Visser, J. (2000). Synergy between enzymes from *Aspergillus* involved in the degradation of plant cell wall polysaccharides. *Carbohydrate Research*, 327(4), 401–410.
- Demarche, P., Junghanns, C., Nair, R. R., & Agathos, S. N. (2011). Harnessing the power of enzymes for environmental stewardship. *Biotechnology Advances*, 30(5), 933–953.
- Demirelli, M., Karaoğlu, E., Baykal, a., Sözeri, H., & Uysal, E. (2014). Synthesis, characterization and catalytic activity of CoFe₂O₄-APTES-Pd magnetic recyclable catalyst. *Journal of Alloys and Compounds*, 582, 201–207.
- Dhiman, S. S., Jagtap, S. S., Jeya, M., Haw, J. R., Kang, Y. C., & Lee, J. K. (2012). Immobilization of Pholiota adiposa xylanase onto SiO₂ nanoparticles and its application for production of xylooligosaccharides. *Biotechnology Letters*, 34(7), 1307–1313.
- Dong, T., Zhao, L., Huang, Y., & Tan, X. (2010). Preparation of cross-linked aggregates of aminoacylase from *Aspergillus melleus* by using bovine serum albumin as an inert additive. *Bioresource Technology*, 101(16), 6569–71.
- Dutta, S. K., & Chakraborty, S. (2015). Kinetic analysis of two-phase enzymatic hydrolysis of hemicellulose of xylan type. *Bioresource Technology*, 198, 642–650.
- Eisenhuber, K., Krennhuber, K., Steinmüller, V., & Jäger, A. (2013). Comparison of Different Pre-treatment Methods for Separating Hemicellulose from Straw During Lignocellulose Bioethanol Production. *Energy Procedia*, 40, 172–181.
- Flesch, C., Joubert, M., Bourgeat-Lami, E., Mornet, S., Duguet, E., Delaite, C., & Dumas, P. (2005). Organosilane-modified maghemite nanoparticles and their use as co-initiator in the ring-opening polymerization of ϵ -caprolactone. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 262(1–3), 150–157.
- Fuzi, S.F.Z.M. (2013). *Enhanced Production of Recombinant β -1,4 Endoxylanase by Kluyveromyces lactis GG799*, PhD Thesis, Universiti Teknologi Malaysia.

- Gan, C.-Y., Alkarkhi, A. F. M., & Easa, A. M. (2009). Using response surface methodology to optimize process parameters and cross-linking agents for production of combined-cross-linked bovine serum albumin gels. *Journal of Bioscience and Bioengineering*, 107(4), 366–72.
- Garcia-Galan, C., Berenguer-Murcia, Á., Fernandez-Lafuente, R., & Rodrigues, R. C. (2011). Potential of Different Enzyme Immobilization Strategies to Improve Enzyme Performance. *Advanced Synthesis & Catalysis*, 353(16), 2885–2904.
- García-García, M. I., Sola-Carvajal, A., Sánchez-Carrón, G., García-Carmona, F., & Sánchez-Ferrer, A. (2011). New stabilized FastPrep-CLEAs for sialic acid synthesis. *Bioresource Technology*, 102(10), 6186–91.
- George, P., Lundin, L., & Kasapis, S. (2014). Thermomechanical effects of co-solute on the structure formation of bovine serum albumin. *Food Chemistry*, 157, 296–301.
- Gokhale, A. a., Lu, J., & Lee, I. (2013). Immobilization of cellulase on magnetoresponsive graphene nano-supports. *Journal of Molecular Catalysis B: Enzymatic*, 90, 76–86.
- Gunda, N. S. K., Singh, M., Norman, L., Kaur, K., & Mitra, S. K. (2014). Optimization and characterization of biomolecule immobilization on silicon substrates using (3-aminopropyl)triethoxysilane (APTES) and glutaraldehyde linker. *Applied Surface Science*, 305, 522–530.
- Gupta, K., Jana, A. K., Kumar, S., & Maiti, M. (2013). Immobilization of amyloglucosidase from SSF of *Aspergillus niger* by cross-linked enzyme aggregate onto magnetic nanoparticles using minimum amount of carrier and characterizations. *Journal of Molecular Catalysis B: Enzymatic*, 98, 30–36.
- He, J., Chen, D., Yu, B., & Zhang, K. (2010). Optimization of the *Trichoderma reesei* endo-1,4-beta-xylanase production by recombinant *Pichia pastoris*. *Biochemical Engineering Journal*, 52(1), 1–6.
- Hormigo, D., García-Hidalgo, J., Acebal, C., de la Mata, I., & Arroyo, M. (2012a). Preparation and characterization of cross-linked enzyme aggregates (CLEAs) of recombinant poly-3-hydroxybutyrate depolymerase from *Streptomyces exfoliatus*. *Bioresource Technology*, 115, 177–82.
- Hu, B., Pan, J., Yu, H. L., Liu, J. W., & Xu, J. H. (2009). Immobilization of *Serratia marcescens* lipase onto amino-functionalized magnetic nanoparticles for repeated use in enzymatic synthesis of Diltiazem intermediate. *Process Biochemistry*.

- Idris, A., Ismail, N. S. M., Hassan, N., Misran, E., & Ngomsik, A. F. (2012). Synthesis of magnetic alginate beads based on maghemite nanoparticles for Pb(II) removal in aqueous solution. *Journal of Industrial and Engineering Chemistry*, 18(5), 1582–1589.
- Illanes, A. (1999). Stability of biocatalysts. *Electronic Journal of Biotechnology*, 2(1), 1–9.
- Islan, G. a., Martinez, Y. N., Illanes, a., & Castro, G. R. (2014). Development of novel alginate lyase cross-linked aggregates for the oral treatment of cystic fibrosis. *RSC Advances*, 4(23), 11758.
- Iyer, P. V., & Ananthanarayan, L. (2008). Enzyme stability and stabilization—Aqueous and non-aqueous environment. *Process Biochemistry*, 43(10), 1019–
- Jafari Khorshidi, K., Lenjannezhadian, H., Jamalan, M., & Zeinali, M. (2015). Preparation and characterization of nanomagnetic cross-linked cellulase aggregates for cellulose bioconversion. *Journal of Chemical Technology & Biotechnology*, 91(2), 539–546.
- Jasti, L. S., Dola, S. R., Kumaraguru, T., Bajja, S., Fadnavis, N. W., Addepally, U., Rajdeo, K., Ponrathnam, S. & Deokar, S. (2014). Protein-coated polymer as a matrix for enzyme immobilization: immobilization of trypsin on bovine serum albumin-coated allyl glycidyl ether-ethylene glycol dimethacrylate copolymer. *Biotechnology Progress*, 30(2), 317–23.
- Ji, C., Nguyen, L. N., Hou, J., Hai, F. I., & Chen, V. (2017). Direct immobilization of laccase on titania nanoparticles from crude enzyme extracts of *P. ostreatus* culture for micro-pollutant degradation. *Separation and Purification Technology*, 178, 215–223.
- Ju, H., Jang, E., Ryu, B. H., & Kim, T. D. (2013). Characterization and preparation of highly stable aggregates of a novel type of hydrolase (BL28) from *Bacillus licheniformis*. *Bioresource Technology*, 128, 81–6.
- Jun, H., Bing, Y., Keying, Z., Xuemei, D., & Daiwen, C. (2009). Expression of *Trichoderma reesei* beta-xylanase gene in *Escherichia coli* and activity of the enzyme on fiber-bound substrates. *Protein Expression & Purification*, 67(1), 1–6.
- Jung, D.-H., Jung, J.-H., Seo, D.-H., Ha, S.-J., Kweon, D.-K., & Park, C.-S. (2013). One-pot bioconversion of sucrose to trehalose using enzymatic sequential reactions in combined cross-linked enzyme aggregates. *Bioresource Technology*, 2011–2014.

- Kang, S. C., Jo, Y. J., Bak, J. P., Kim, K.-C., & Kim, Y.-S. (2007). Evaluation for protein binding affinity of maghemite and magnetite nanoparticles. *Journal of Nanoscience and Nanotechnology*, 7 (11), 3706-8.
- Kartal, F., Janssen, M. H. A., Hollmann, F., Sheldon, R. A., & Kılinc, A. (2011). Improved esterification activity of *Candida rugosa* lipase in organic solvent by immobilization as Cross-linked enzyme aggregates (CLEAs). *Journal of Molecular Catalysis. B, Enzymatic*, 71(3–4), 85–89.
- Kartal, F., & Kılinc, A. (2012). Cross-linked aggregates of *Rhizopus oryzae* lipase as industrial biocatalysts: Preparation, optimization, characterization, and application for enantioselective resolution reactions. *Biotechnology Progress*, 28(4), 937–945.
- Khanahmadi, S., Yusof, F., Amid, A., Mahmud, S. S., & Mahat, M. K. (2015). Optimized preparation and characterization of CLEA-lipase from cocoa pod husk. *Journal of Biotechnology*, 202, 153–161.
- Kim, I. J., Youn, H. J., & Kim, K. H. (2016). Synergism of an auxiliary activity 9 (AA9) from *Chaetomium globosum* with xylanase on the hydrolysis of xylan and lignocellulose. *Process Biochemistry*, 51(10), 1445–1451.
- Kim, M. H., Park, S., Kim, Y. H., Won, K., & Lee, S. H. (2013). Immobilization of formate dehydrogenase from *Candida boidinii* through cross-linked enzyme aggregates. *Journal of Molecular Catalysis B: Enzymatic*, 97, 209–214.
- Kisonen, V., Xu, C., Eklund, P., Lindqvist, H., Sundberg, A., Pranovich, A., Sinkkonen, J., Vilaplana, F. & Willför, S. (2014). Cationised O-acetyl galactoglucomannans: Synthesis and characterisation. *Carbohydrate Polymers*, 99, 755–764.
- Kluchova, K., Zboril, R., Tucek, J., Pecova, M., Zajoncova, L., Safarik, I., Jancik, D., Bartonkova, H., Bellesi, V., Novak, P. & Petridis, D. (2009). Superparamagnetic maghemite nanoparticles from solid-state synthesis - their functionalization towards peroral MRI contrast agent and magnetic carrier for trypsin immobilization. *Biomaterials*, 30(15), 2855–63.
- Kopp, W., da Costa, T. P., Pereira, S. C., Jafelicci Jr, M., Giordano, R. C., Marques, R. F. C., Araújo-Moreira, F.M. & Giordano, R. L. C. (2013). Easily handling penicillin G acylase magnetic cross-linked enzymes aggregates: Catalytic and morphological studies. *Process Biochemistry*, 49(1), 38–46.

- Krajewska, B. (2004). Application of chitin- and chitosan-based materials for enzyme immobilizations : a review, 35, 126–139.
- Kumar, P., Barrett, D. M., Delwiche, M. J., & Stroeve, P. (2009). Methods for Pretreatment of Lignocellulosic Biomass for Efficient Hydrolysis and Biofuel Production. *Industrial & Engineering Chemistry Research*, 48(8), 3713–3729.
- Kumar, S., Mohan, U., Kamble, A. L., Pawar, S., & Banerjee, U. C. (2010). Cross-linked enzyme aggregates of recombinant *Pseudomonas putida* nitrilase for enantioselective nitrile hydrolysis. *Bioresource Technology*, 101(17), 6856–8.
- Kumar, V. V., Sivanesan, S., & Cabana, H. (2014). Magnetic cross-linked laccase aggregates - Bioremediation tool for decolorization of distinct classes of recalcitrant dyes. *The Science of the Total Environment*, 487, 830–9.
- Kurtan, U., & Baykal, A. (2014). Fabrication and characterization of Fe₃O₄@APTES@PAMAM-Ag highly active and recyclable magnetic nanocatalyst: Catalytic reduction of 4-nitrophenol. *Materials Research Bulletin*, 60, 79–87.
- Laane, C., Boeren, S., Vos, K., & Veeger, C. (1987). Rules for optimization of biocatalysis in organic solvents. *Biotechnology and Bioengineering*, 30(1), 81–87.
- Laemmli, U. K. (1970). Cleavage of Structural Proteins during the Assembly of the Head of Bacteriophage T4. *Nature*, 227, 680-685.
- Lai, J.-Q., Hu, Z.-L., Sheldon, R. a., & Yang, Z. (2012). Catalytic performance of cross-linked enzyme aggregates of *Penicillium expansum* lipase and their use as catalyst for biodiesel production. *Process Biochemistry*, 47, 2058–2063.
- Li, B., Dong, S. L., Xie, X. L., Xu, Z. B., & Li, L. (2012). Preparation and Properties of Cross-Linked Enzyme Aggregates of Cellulase. *Advanced Materials Research*, 581–582, 257–260.
- Li, D., Teoh, W. Y., Gooding, J. J., Selomulya, C., & Amal, R. (2010). Functionalization Strategies for Protease Immobilization on Magnetic Nanoparticles. *Advanced Functional Materials*, 20(11), 1767–1777.
- Li, L., Li, G., Cao, L., Ren, G., Kong, W., Wang, S., Guo, G. S. & Liu, Y. H. (2015). Characterization of the Cross-Linked Enzyme Aggregates of a Novel β -Galactosidase, a Potential Catalyst for the Synthesis of Galacto-Oligosaccharides. *Journal of Agricultural and Food Chemistry*, 63(3), 894–901.

- Limayem, A., & Ricke, S. C. (2012). Lignocellulosic biomass for bioethanol production: Current perspectives, potential issues and future prospects. *Progress in Energy and Combustion Science*, 38(4), 449–467.
- Lin, Q., Li, H., Ren, J., Deng, A., Li, W., & Liu, C. (2017). Production of xylooligosaccharides by microwave-induced , organic acid-catalyzed hydrolysis of different xylan-type hemicelluloses : Optimization by response surface methodology. *Carbohydrate Polymers*, 157, 214–225.
- Lineweaver, H., & Burk, D. (1934). The determination of enzyme dissociation constants. *Journal of the American Chemical Society*, 56(3), 658–666.
- Liu, N., Liang, G., Dong, X., Qi, X., Kim, J., & Piao, Y. (2016). Stabilized magnetic enzyme aggregates on graphene oxide for high performance phenol and bisphenol A removal. *Chemical Engineering Journal*, 306, 1026–1034.
- López-Gallego, F., Betancor, L., Hidalgo, A., Alonso, N., Fernández-Lafuente, R., & Guisán, J. M. (2005). Co-aggregation of enzymes and polyethyleneimine: a simple method to prepare stable and immobilized derivatives of glutaryl acylase. *Biomacromolecules*, 6(4), 1839–42.
- Mahmod, S. S., Yusof, F., Jami, M. S., & Khanahmadi, S. (2016). Optimizing the preparation conditions and characterization of a stable and recyclable cross-linked enzyme aggregate (CLEA)-protease. *Bioresources and Bioprocessing*, 3(1), 3.
- Mahmod, S. S., Yusof, F., Jami, M. S., Khanahmadi, S., & Shah, H. (2015). Development of an immobilized biocatalyst with lipase and protease activities as a multipurpose cross-linked enzyme aggregate (multi-CLEA). *Process Biochemistry*, 50(12), 2144–2157.
- Maitan-Alfenas, G. P., Oliveira, M. B., Nagem, R. A. P., de Vries, R. P., & Guimarães, V. M. (2016). Characterization and biotechnological application of recombinant xylanases from *Aspergillus nidulans*. *International Journal of Biological Macromolecules*, 91, 60–67.
- Manju, S., & Singh Chadha, B. (2011). Production of hemicellulolytic enzymes for hydrolysis of lignocellulosic biomass. *Biofuels* (1st ed.). Elsevier Inc.
- Martin-Sampedro, R., Eugenio, M. E., Moreno, J. A., Revilla, E., & Villar, J. C. (2014). Integration of a kraft pulping mill into a forest biorefinery: Pre-extraction of hemicellulose by steam explosion versus steam treatment. *Bioresource Technology*, 153, 236–244.

- Martin-Sampedro, R., Revilla, E., Villar, J. C., & Eugenio, M. E. (2014). Enhancement of enzymatic saccharification of *Eucalyptus globulus*: Steam explosion versus steam treatment. *Bioresource Technology*, 167, 186–191.
- Massart, R. (1981). Preparation of aqueous magnetic liquids in alkaline and acidic media. *IEEE Transactions on Magnetics*.
- Mateo, C., Palomo, J. M., van Langen, L. M., van Rantwijk, F., & Sheldon, R. a. (2004). A new, mild cross-linking methodology to prepare cross-linked enzyme aggregates. *Biotechnology and Bioengineering*, 86(3), 273–6.
- Menon, V., & Rao, M. (2012). Trends in bioconversion of lignocellulose: Biofuels, platform chemicals & biorefinery concept. *Progress in Energy and Combustion Science*, 38(4), 522–550.
- Migneault, I., Dartiguenave, C., Bertrand, M. J., & Waldron, K. C. (2004). Glutaraldehyde: behavior in aqueous solution, reaction with proteins, and application to enzyme cross-linking. *BioTechniques*, 37(5), 790–6, 798–802.
- Miller, G. L. (1959). Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. *Analytical Chemistry*, 31(3), 426–428.
- Min, K., Kim, J., Park, K., & Yoo, Y. J. (2012). Enzyme immobilization on carbon nanomaterials: Loading density investigation and zeta potential analysis. *Journal of Molecular Catalysis B: Enzymatic*, 83, 87–93.
- Montoro-García, S., Gil-Ortiz, F., Navarro-Fernández, J., Rubio, V., García-Carmona, F., & Sánchez-Ferrer, A. (2010). Improved cross-linked enzyme aggregates for the production of desacetyl beta-lactam antibiotics intermediates. *Bioresource Technology*, 101(1), 331–6.
- Moure, A., Gullón, P., Domínguez, H., & Parajó, J. C. (2006). Advances in the manufacture, purification and applications of xylo-oligosaccharides as food additives and nutraceuticals. *Process Biochemistry*, 41(9), 1913–1923.
- Nadar, S. S., Muley, A. B., Ladole, M. R., & Joshi, P. U. (2016). Macromolecular cross-linked enzyme aggregates (M-CLEAs) of α -amylase. *International Journal of Biological Macromolecules*, 84, 69–78.
- Nadar, S. S., & Rathod, V. K. (2016). Magnetic macromolecular cross linked enzyme aggregates (CLEAs) of glucoamylase. *Enzyme and Microbial Technology*, 83, 78–87.

- Nagar, S., Mittal, A., Kumar, D., Kumar, L., & Gupta, V. K. (2012). Immobilization of xylanase on glutaraldehyde activated aluminum oxide pellets for increasing digestibility of poultry feed. *Process Biochemistry*, 47(9), 1402–1410.
- Nakazawa, H., Kawai, T., Ida, N., Shida, Y., Shioya, K., Kobayashi, Y., Okada, H., Tani, S., Sumitani, J.I., Kawaguchi, T., Morikawa, Y. & Ogasawara, W. (2016). A high performance *Trichoderma reesei* strain that reveals the importance of xylanase III in cellulosic biomass conversion. *Enzyme and Microbial Technology*, 82, 89–95.
- Nath, A., & Chattopadhyay, P. K. (2007). Optimization of oven toasting for improving crispness and other quality attributes of ready to eat potato-soy snack using response surface methodology. *Journal of Food Engineering*, 80(4), 1282–1292.
- Negahdar, L., Delidovich, I., & Palkovits, R. (2016). Aqueous-phase hydrolysis of cellulose and hemicelluloses over molecular acidic catalysts: Insights into the kinetics and reaction mechanism. *Applied Catalysis B: Environmental*, 184, 285–298.
- Netto, C. G. C. M., Toma, H. E., & Andrade, L. H. (2013). Superparamagnetic nanoparticles as versatile carriers and supporting materials for enzymes. *Journal of Molecular Catalysis B: Enzymatic*, 85–86, 71–92.
- Nguyen, L. T., Seow, N., & Yang, K. L. (2017). Hollow cross-linked enzyme aggregates (h-CLEA) of laccase with high uniformity and activity. *Colloids and Surfaces B: Biointerfaces*, 151, 88–94.
- Nguyen, L. T., & Yang, K. L. (2017). Combined cross-linked enzyme aggregates of horseradish peroxidase and glucose oxidase for catalyzing cascade chemical reactions. *Enzyme and Microbial Technology*, 100, 52–59.
- Olcay, H., Subrahmanyam, A. V., Xing, R., Lajoie, J., Dumesic, J. A., & Huber, G. W. (2013). Production of renewable petroleum refinery diesel and jet fuel feedstocks from hemicellulose sugar streams. *Energy Environ. Sci.*
- Otieno, D. O., & Ahring, B. K. (2012). The potential for oligosaccharide production from the hemicellulose fraction of biomasses through pretreatment processes: Xylooligosaccharides (XOS), arabinooligosaccharides (AOS), and mannoooligosaccharides (MOS). *Carbohydrate Research*, 360, 84–92.

- Pan, J., Kong, X.-D., Li, C.-X., Ye, Q., Xu, J.-H., & Imanaka, T. (2011). Cross-linking of enzyme coaggregate with polyethyleneimine: A simple and promising method for preparing stable biocatalyst of *Serratia marcescens* lipase. *Journal of Molecular Catalysis B: Enzymatic*, 68(3–4), 256–261.
- Patel, H., Chapla, D., & Shah, A. (2017). Bioconversion of pretreated sugarcane bagasse using enzymatic and acid followed by enzymatic hydrolysis approaches for bioethanol production. *Renewable Energy*, 109, 323–331.
- Peng, F., Peng, P., Xu, F., & Sun, R.-C. (2012). Fractional purification and bioconversion of hemicelluloses. *Biotechnology Advances*, 30(4), 879–903.
- Peng, P., & She, D. (2014). Isolation, structural characterization, and potential applications of hemicelluloses from bamboo: A review. *Carbohydrate Polymers*, 112, 701–720.
- Pointing, S. B. (1999). Qualitative methods for the determination of lignocellulolytic enzyme production by tropical fungi. *Fungal Diverssity*, 2(March), 17–33.
- Polizeli, M. L. T. M., Rizzatti, a C. S., Monti, R., Terenzi, H. F., Jorge, J. a, & Amorim, D. S. (2005). Xylanases from fungi: properties and industrial applications. *Applied Microbiology and Biotechnology*, 67(5), 577–91.
- Rao, M. A., Scelza, R., Acevedo, F., Diez, M. C., & Gianfreda, L. (2014). Chemosphere Enzymes as useful tools for environmental purposes. *Chemosphere*, 107, 145–162.
- Reza, R. T., Martínez Pérez, C. a., Rodríguez González, C. a., Romero, H. M., & García Casillas, P. E. (2010). Effect of the polymeric coating over Fe₃O₄ particles used for magnetic separation. *Central European Journal of Chemistry*.
- Rissanen, J. V., Murzin, D. Y., Salmi, T., & Grénman, H. (2015). Aqueous extraction of hemicelluloses from spruce - From hot to warm. *Bioresource Technology*, 199, 279–282.
- Rodrigues, R. C., Barbosa, O., Ortiz, C., Berenguer-Murcia, Á., Torres, R., & Fernandez-Lafuente, R. (2014). Amination of enzymes to improve biocatalyst performance: coupling genetic modification and physicochemical tools. *RSC Advances*, 4(72), 38350.
- Royvaran, M., Taheri-Kafrani, A., Landarani Isfahani, A., & Mohammadi, S. (2016). Functionalized superparamagnetic graphene oxide nanosheet in enzyme engineering: A highly dispersive, stable and robust biocatalyst. *Chemical Engineering Journal*, 288, 414–422.

- Sabiha-Hanim, S., Noor, M. A. M., & Rosma, A. (2011). Effect of autohydrolysis and enzymatic treatment on oil palm (*Elaeis guineensis* Jacq.) frond fibres for xylose and xylooligosaccharides production. *Bioresource Technology*, 102(2), 1234–1239.
- Sakamoto, Y., Blanco, R. M., & Di, I. (2008). Immobilization of lipase in ordered mesoporous materials : Effect of textural and structural parameters, 114, 201–213.
- Samanta, a. K., Jayapal, N., Kolte, a. P., Senani, S., Sridhar, M., Suresh, K. P., & Sampath, K. T. (2012). Enzymatic production of xylooligosaccharides from alkali solubilized xylan of natural grass (*Sehima nervosum*). *Bioresource Technology*, 112, 199–205.
- Sanchez, S., & Demain, A. L. (2011). Enzymes and Bioconversions of Industrial , Pharmaceutical , and Biotechnological Abstract :, 15(1), 224–230.
- Sassolas, A., Blum, L. J., & Leca-Bouvier, B. D. (2012). Immobilization strategies to develop enzymatic biosensors. *Biotechnology Advances*, 30(3), 489–511.
- Schoe, Wvaart, Rolbers, M. W., Golubovic, M., Ottens, M., Kieboom, a P. G., van Rantwijk, F., van der Wielen, L.A. & Sheldon, R. a. (2004). Preparation, optimization, and structures of cross-linked enzyme aggregates (CLEAs). *Biotechnology and Bioengineering*, 87(6), 754–62.
- Shah, S., Sharma, A., & Gupta, M. N. (2006). Preparation of cross-linked enzyme aggregates by using bovine serum albumin as a proteic feeder. *Analytical Biochemistry*, 351(2), 207–13.
- Shahrestani, H., Taheri-Kafrani, A., Soozanipour, A., & Tavakoli, O. (2016). Enzymatic clarification of fruit juices using xylanase immobilized on 1,3,5-triazine-functionalized silica-encapsulated magnetic nanoparticles. *Biochemical Engineering Journal*, 109, 51–58.
- Sheldon, R. a. (2007). Cross-linked enzyme aggregates (CLEAs): stable and recyclable biocatalysts. *Biochemical Society Transactions*, 35(Pt 6), 1583–7.
- Sheldon, R. a. (2011). Characteristic features and biotechnological applications of cross-linked enzyme aggregates (CLEAs). *Applied Microbiology and Biotechnology*, 92(3), 467–77.
- Sheldon, R. A. (2007). Enzyme Immobilization: The Quest for Optimum Performance. *Advanced Synthesis & Catalysis*, 349(8–9), 1289–1307.

- Sheldon, R. A. (2011). Cross-Linked Enzyme Aggregates as Industrial Biocatalysts Abstract :, 213(1), 213–223.
- Shengping Lia, Yintao Sua, Yadan Liua, Lifang Suna, Minxiang Yub, Y. W. (2016). Preparation and characterization of cross-linked enzyme aggregates (CLEAs) of recombinant thermostable alkylsulfatase (SdsAP) from. *Process Biochemistry*, 51(12), 2084–2089.
- Shin, Y.-O., Wahnou, D., Weber, M. E., & Vera, J. H. (2004). Selective precipitation and recovery of xylanase using surfactant and organic solvent. *Biotechnology and Bioengineering*, 86(6), 698–705.
- Silva-Fernandes, T., Duarte, L. C., Carneiro, F., Marques, S., Loureiro-Dias, M. C., Fonseca, C., & Gírio, F. (2015). Biorefining strategy for maximal monosaccharide recovery from three different feedstocks: eucalyptus residues, wheat straw and olive tree pruning. *Bioresource Technology*, 183, 203–212.
- Singh, R., Kumar, M., Mittal, A., & Mehta, P. K. (2016). Microbial enzymes: industrial progress in 21st century. 3 *Biotech*, 6(2), 1–15.
- Soozanipour, A., Taheri-Kafrani, A., & Landarani Isfahani, A. (2015). Covalent attachment of xylanase on functionalized magnetic nanoparticles and determination of its activity and stability. *Chemical Engineering Journal*, 270, 235–243.
- Souillac, P. O., Costantino, H. R., Middaugh, C. R., & Rytting, J. H. (2002). Investigation of Protein / Carbohydrate Interactions in the Dried State . 1 . Calorimetric Studies. *International Journal of Pharmaceutics*, 91 (1), 206-216.
- Stressler, T., Ewert, J., Eisele, T., & Fischer, L. (2015). Cross-linked enzyme aggregates (CLEAs) of PepX and PepN - production, partial characterization and application of combi-CLEAs for milk protein hydrolysis. *Biocatalysis and Agricultural Biotechnology*, 4(4), 752–760.
- Šulek, F., Drofenik, M., Habulin, M., & Knez, Ž. (2010). Surface functionalization of silica-coated magnetic nanoparticles for covalent attachment of cholesterol oxidase. *Journal of Magnetism and Magnetic Materials*, 322(2), 179–185.
- Šulek, F., Knez, Ž., & Habulin, M. (2010). Immobilization of cholesterol oxidase to finely dispersed silica-coated maghemite nanoparticles based magnetic fluid. *Applied Surface Science*, 256(14), 4596–4600.

- Sun, W. Q., & Davidson, P. (1998). Protein inactivation in amorphous sucrose and trehalose matrices: Effects of phase separation and crystallization. *Biochimica et Biophysica Acta - General Subjects*, 1425(1), 235-44.
- Taboada-Puig, R., Junghanns, C., Demarche, P., Moreira, M. T., Feijoo, G., Lema, J. M., & Agathos, S. N. (2011). Combined cross-linked enzyme aggregates from versatile peroxidase and glucose oxidase: production, partial characterization and application for the elimination of endocrine disruptors. *Bioresource Technology*, 102(11), 6593–9.
- Talekar, S., Desai, S., Pillai, M., Nagavekar, N., Ambarkar, S., Surnis, S., Ladole, M., Nadar, S. & Mulla, M. (2013). Carrier free co-immobilization of glucoamylase and pullulanase as combi-cross linked enzyme aggregates (combi-CLEAs). *RSC Advances*, 3(7), 2265.
- Talekar, S., Ghodake, V., Ghotage, T., Rathod, P., Deshmukh, P., Nadar, S., Mulla, M. & Ladole, M. (2012). Novel magnetic cross-linked enzyme aggregates (magnetic CLEAs) of alpha amylase. *Bioresource Technology*, 123, 542–547.
- Talekar, S., Joshi, A., Joshi, G., Kamat, P., Haripurkar, R., & Kambale, S. (2013). Parameters in preparation and characterization of cross-linked enzyme aggregates (CLEA). *RSC Advances*, 3(31), 12485-12511.
- Talekar, S., Joshi, A., Chougale, R., Nakhe, A., & Bhojwani, R. (2016). Immobilized enzyme mediated synthesis of silver nanoparticles using cross-linked enzyme aggregates (CLEAs) of NADH-dependent nitrate reductase. *Nano-Structures & Nano-Objects*, 6(April), 23–33.
- Talekar, S., Joshi, G., Chougale, R., Nainegali, B., Desai, S., Joshi, A., Kambale, S., Kamat, P., Haripurkar, R., Jadhav, S. & Nadar, S. (2014). Preparation of stable cross-linked enzyme aggregates (CLEAs) of NADH-dependent nitrate reductase and its use for silver nanoparticle synthesis from silver nitrate. *Catalysis Communications*, 53, 62–66.
- Talekar, S., Nadar, S., Joshi, A., & Joshi, G. (2014). Pectin cross-linked enzyme aggregates (pectin-CLEAs) of glucoamylase. *RSC Advances*, 4(103), 59444–53.
- Talekar, S., Shah, V., Patil, S., & Nimbalkar, M. (2012). Porous cross linked enzyme aggregates (p-CLEAs) of *Saccharomyces cerevisiae* invertase. *Catalysis Science & Technology*, 2(8), 1575.

- Terrasan, C. R. F., Guisan, J. M., & Carmona, E. C. (2016). Xylanase and β -xylosidase from *Penicillium janczewskii*: Purification, characterization and hydrolysis of substrates. *Electronic Journal of Biotechnology*, 23, 54–62.
- Torabizadeh, H., Tavakoli, M., & Safari, M. (2014). Immobilization of thermostable α -amylase from *Bacillus licheniformis* by cross-linked enzyme aggregates method using calcium and sodium ions as additives. *Journal of Molecular Catalysis B: Enzymatic*, 108, 13–20.
- Törrönen, A., & Rouvinen, J. (1995). Structural Comparison of Two Major endo-1,4-Xylanases from *Trichoderma reesei*. *Biochemistry*, 34 (3), 847–56.
- Tscheliessnig, A., Satzer, P., Hammerschmidt, N., Schulz, H., Helk, B., & Jungbauer, A. (2014). Ethanol precipitation for purification of recombinant antibodies. *Journal of Biotechnology*, 188C, 17–28.
- Tudorache, M., Nae, A., Coman, S., & Parvulescu, V. I. (2013). Strategy of cross-linked enzyme aggregates onto magnetic particles adapted to the green design of biocatalytic synthesis of glycerol carbonate. *RSC Advances*, 3(12), 4052.
- Tükel, S. S., Hürrem, F., Yildirim, D., & Alptekin, Ö. (2013). Preparation of cross-linked enzyme aggregates (CLEA) of catalase and its characterization. *Journal of Molecular Catalysis B: Enzymatic*, 97, 252–257.
- Uçkun Kiran, E., Akpınar, O., & Bakır, U. (2013). Improvement of enzymatic xylooligosaccharides production by the co-utilization of xylans from different origins. *Food and Bioproducts Processing*, 91(4), 565–574.
- Vafiadi, C., Topakas, E., & Christakopoulos, P. (2008). Preparation of multipurpose cross-linked enzyme aggregates and their application to production of alkyl ferulates. *Journal of Molecular Catalysis B: Enzymatic*, 54(1–2), 35–41.
- Vaidya, B. K., Kuwar, S. S., Golegaonkar, S. B., & Nene, S. N. (2012). Preparation of cross-linked enzyme aggregates of l-aminoacylase via co-aggregation with polyethyleneimine. *Journal of Molecular Catalysis B: Enzymatic*, 74(3–4), 184–191.
- Valdés, E. C., Soto, L. W., & Arcaya, G. A. (2011). Influence of the pH of glutaraldehyde and the use of dextran aldehyde on the preparation of cross-linked enzyme aggregates (CLEAs) of lipase from *Burkholderia cepacia*. *Electronic Journal of Biotechnology*, 14(3), 1–7.

- Van Dyk, J. S., & Pletschke, B. I. (2012). A review of lignocellulose bioconversion using enzymatic hydrolysis and synergistic cooperation between enzymes-Factors affecting enzymes, conversion and synergy. *Biotechnology Advances*, 30(6), 1458–80.
- van Gool, M. P., Toth, K., Schols, H. a, Szakacs, G., & Gruppen, H. (2012). Performance of hemicellulolytic enzymes in culture supernatants from a wide range of fungi on insoluble wheat straw and corn fiber fractions. *Bioresource Technology*, 114, 523–8.
- Velmurugan, R., & Incharoensakdi, A. (2017). MgO-Fe₃O₄ linked cellulase enzyme complex improves the hydrolysis of cellulose from *Chlorella sp.* CYB2. *Biochemical Engineering Journal*, 122, 22–30.
- Vinoth, V., Sivanesan, S., & Cabana, H. (2014). Science of the Total Environment Magnetic cross-linked laccase aggregates — Bioremediation tool for decolorization of distinct classes of recalcitrant dyes, 487, 830–839.
- Wan Azelee, N. I. (2015). *Optimisation of Kenaf Pretreatment and Enzymatic Reaction Conditions of Kenaf Hemicellulose for xylooligosaccharides Production*. PhD Thesis, Universiti Teknologi Malaysia.
- Wang, A., Zhang, F., Chen, F., Wang, M., Li, H., Zeng, Z., Xie, T. & Chen, Z. (2011). A facile technique to prepare cross-linked enzyme aggregates using p-benzoquinone as cross-linking agent. *Korean Journal of Chemical Engineering*, 28(4), 1090–1095.
- Wang, M., Jia, C., Qi, W., Yu, Q., Peng, X., Su, R., & He, Z. (2011). Porous-CLEAs of papain: application to enzymatic hydrolysis of macromolecules. *Bioresource Technology*, 102(3), 3541–5.
- Wang, M., Qi, W., Jia, C., Ren, Y., Su, R., & He, Z. (2011). Enhancement of activity of cross-linked enzyme aggregates by a sugar-assisted precipitation strategy: technical development and molecular mechanism. *Journal of Biotechnology*, 156(1), 30–8.
- Wang, P. (2006). Nanoscale biocatalyst systems. *Current Opinion in Biotechnology*, 17(6), 574–579.
- Wang, S., Meng, X., Zhou, H., Liu, Y., Secundo, F., & Liu, Y. (2016). Enzyme Stability and Activity in Non-Aqueous Reaction Systems: A Mini Review. *Catalysts*, 32, 1–16.

- Wang, S., Zheng, D., Yin, L., & Wang, F. (2017). Preparation, activity and structure of cross-linked enzyme aggregates (CLEAs) with nanoparticle. *Enzyme and Microbial Technology*, 107(January), 22–31.
- Wilson, L., Illanes, A., Abián, O., Pessela, B. C. C., Fernández-Lafuente, R., & Guisán, J. M. (2004). Co-aggregation of penicillin G acylase and polyionic polymers: an easy methodology to prepare enzyme biocatalysts stable in organic media. *Biomacromolecules*, 5(3), 852–7.
- Wong, W. H., Lee, W. X., Ramanan, R. N., Tee, L. H., Kong, K. W., Galanakis, C. M., ... Prasad, K. N. (2015). Two level half factorial design for the extraction of phenolics, flavonoids and antioxidants recovery from palm kernel by-product. *Industrial Crops and Products*, 63, 238–248.
- Xu, D.-Y., Yang, Y., & Yang, Z. (2011). Activity and stability of cross-linked tyrosinase aggregates in aqueous and nonaqueous media. *Journal of Biotechnology*, 152(1–2), 30–6.
- Yamaguchi, H., Miyazaki, M., Asanomi, Y., & Maeda, H. (2011). Poly-lysine supported cross-linked enzyme aggregates with efficient enzymatic activity and high operational stability. *Catalysis Science & Technology*, 1(7), 1256.
- Yamaura, M., Camilo, R. ., Sampaio, L. ., Macêdo, M. ., Nakamura, M., & Toma, H. . (2004). Preparation and characterization of (3-aminopropyl)triethoxysilane-coated magnetite nanoparticles. *Journal of Magnetism and Magnetic Materials*, 279(2–3), 210–217.
- Yan, X., Wang, X., Zhao, P., Zhang, Y., Xu, P., & Ding, Y. (2012). Xylanase immobilized nanoporous gold as a highly active and stable biocatalyst. *Microporous and Mesoporous Materials*, 161, 1–6.
- Yang, X., Zheng, P., Ni, Y., & Sun, Z. (2012). Highly efficient biosynthesis of sucrose-6-acetate with cross-linked aggregates of Lipozyme TL 100 L. *Journal of Biotechnology*, 161(1), 27–33.
- Yang, Z., Wu, D., Chen, C., Cheong, K., Deng, Y., Chen, L., Han, B., Chen, N., Zhao, J. & Li, S. (2016). Preparation of xylooligosaccharides from xylan by controlled acid hydrolysis and fast protein liquid chromatography coupled with refractive index detection. *Separation and Purification Technology*, 171, 151–156.
- Yemiş, O., & Mazza, G. (2012). Optimization of furfural and 5-hydroxymethylfurfural production from wheat straw by a microwave-assisted process. *Bioresource Technology*, 109, 215–223.

- Yu, C.-Y., Li, X.-F., Lou, W.-Y., & Zong, M.-H. (2013). Cross-linked enzyme aggregates of Mung bean epoxide hydrolases: a highly active, stable and recyclable biocatalyst for asymmetric hydrolysis of epoxides. *Journal of Biotechnology*, 166(1–2), 12–9.
- Zhang, P., Dong, S.-J., Ma, H.-H., Zhang, B.-X., Wang, Y.-F., & Hu, X.-M. (2015). Fractionation of corn stover into cellulose, hemicellulose and lignin using a series of ionic liquids. *Industrial Crops & Products*, 76, 688–696.
- Zhang, W., Yang, X., Jia, J., Wang, N., Hu, C., & Yu, X. (2015). Surfactant-activated magnetic cross-linked enzyme aggregates (magnetic CLEAs) of *Thermomyces lanuginosus* lipase for biodiesel production. *Journal of Molecular Catalysis. B, Enzymatic*, 115, 83–89.
- Zhang, Z., Donaldson, A. a, & Ma, X. (2012). Advancements and future directions in enzyme technology for biomass conversion. *Biotechnology Advances*, 30(4), 913–9.
- Zhen, Q., Wang, M., Qi, W., Su, R., & He, Z. (2013). Preparation of β -mannanase CLEAs using macromolecular cross-linkers. *Catalysis Science & Technology*, 3(8), 1937.